

Redox regulation of heat shock protein expression in aging and neurodegenerative disorders associated with oxidative stress: A nutritional approach

Review Article

V. Calabrese¹, G. Scapagnini¹, C. Colombrita¹, A. Ravagna¹, G. Pennisi², A. M. Giuffrida Stella¹, F. Galli³, and D. A. Butterfield⁴

Received September 10, 2003 Accepted September 21, 2003 Published online November 7, 2003; © Springer-Verlag 2003

Summary. Oxidative stress has been implicated in mechanisms leading to neuronal cell injury in various pathological states of the brain. Alzheimer's disease (AD) is a progressive disorder with cognitive and memory decline, speech loss, personality changes and synapse loss. Many approaches have been undertaken to understand AD, but the heterogeneity of the etiologic factors makes it difficult to define the clinically most important factor determining the onset and progression of the disease. However, increasing evidence indicates that factors such as oxidative stress and disturbed protein metabolism and their interaction in a vicious cycle are central to AD pathogenesis.

Brains of AD patients undergo many changes, such as disruption of protein synthesis and degradation, classically associated with the heat shock response, which is one form of stress response. Heat shock proteins are proteins serving as molecular chaperones involved in the protection of cells from various forms of stress.

Recently, the involvement of the heme oxygenase (HO) pathway in antidegenerative mechanisms operating in AD has received considerable attention, as it has been demonstrated that the expression of HO is closely related to that of amyloid precursor protein (APP). HO induction occurs together with the induction of other HSPs during various physiopathological conditions. The vasoactive molecule carbon monoxide and the potent antioxidant bilirubin, products of HO-catalyzed reaction, represent a protective system potentially active against brain oxidative injury. Given the broad cytoprotective properties of the heat shock response there is now strong interest in discovering and developing pharmacological agents capable of inducing the heat shock response.

Increasing interest has been focused on identifying dietary compounds that can inhibit, retard or reverse the multi-stage pathophysiological events underlying AD pathology. Alzheimer's disease, in fact, involves a chronic inflammatory response associated with both brain injury and β -amyloid associated pathology. All of the above evidence suggests that stimulation

of various repair pathways by mild stress has significant effects on delaying the onset of various age-associated alterations in cells, tissues and organisms. Spice and herbs contain phenolic substances with potent antioxidative and chemopreventive properties, and it is generally assumed that the phenol moiety is responsible for the antioxidant activity. In particular, curcumin, a powerful antioxidant derived from the curry spice turmeric, has emerged as a strong inducer of the heat shock response. In light of this finding, curcumin supplementation has been recently considered as an alternative, nutritional approach to reduce oxidative damage and amyloid pathology associated with AD. Here we review the importance of the heme oxygenase pathway in brain stress tolerance and its significance as an antidegenerative mechanism potentially important in AD pathogenesis. These findings have offered new perspectives in medicine and pharmacology, as molecules inducing this defense mechanism appear to be possible candidates for novel cytoprotective strategies. In particular, manipulation of endogenous cellular defense mechanisms such as the heat shock response, through nutritional antioxidants or pharmacological compounds, represents an innovative approach to therapeutic intervention in diseases causing tissue damage, such as neurodegeneration. Consistent with this notion, maintenance or recovery of the activity of vitagenes, such as the HO gene, conceivably may delay the aging process and decrease the occurrence of age-related neurodegenerative diseases.

Keywords: Oxidative stress – AD pathogenesis – Heme oxygenase pathway – Vitamin E – Vitamin C – Micronutrients – Antioxidants

The heat shock pathway of cell stress tolerance

Living cells are continually challenged by conditions which cause acute or chronic stress. To adapt to environmental

¹ Department of Chemistry, Section of Biochemistry and Molecular Biology, Faculty of Medicine, University of Catania, Catania, Italy

² Department of Neurological Sciences, Faculty of Medicine, University of Catania, Catania, Italy

³ Section of Clinical Biochemistry, DI.M.I., University of Perugia, Perugia, Italy

⁴ Department of Chemistry, Center of Membrane Sciences, and Sanders-Brown Center on Aging, University of Kentucky, Lexington, Kentucky, U.S.A.

V. Calabrese et al.

changes and survive different types of injuries, eukaryotic cells have evolved networks of different responses which detect and control diverse forms of stress. One of these responses, known as the heat shock response, has attracted a great deal of attention as a universal fundamental mechanism necessary for cell survival under a wide variety of toxic conditions. In mammalian cells heat shock protein (HSP) synthesis is induced not only after hyperthermia, but also following alterations in the intracellular redox environment, exposure to heavy metals, amino acid analogs or cytotoxic drugs. While prolonged exposure to conditions of extreme stress is harmful and can lead to cell death, induction of HSP synthesis can result in stress tolerance and cytoprotection against stress-induced molecular damage. Furthermore, transient exposure to elevated temperatures has a cross-protective effect against sustained, normally lethal exposures to other pathogenic stimuli. Hence, the heat shock response contributes to establish a cytoprotective state in a variety of metabolic disturbances and injuries, including stroke, epilepsy, cell and tissue trauma, neurodegenerative disease and aging (Mattson et al., 2002; Calabrese et al., 2001a). This has opened new perspectives in medicine and pharmacology, as molecules activating this defense mechanism appear as possible candidates for novel cytoprotective strategies (Calabrese et al., 1998, 2001b, 2002a; Wheller et al., 2003; Misseri et al., 2003; Ropeleski et al., 2003; Goodman and Blank, 2002). In mammalian cells the induction of the heat shock response requires the activation and translocation to the nucleus of one or more heat shock transcription factors which control the expression of a specific set of genes encoding cytoprotective heat shock proteins. Some of the known HSPs include ubiquitin, HSP10, HSP27, HSP32 (or HO-1), HSP47, HSP60, HSC70, HSP70 (or HSP72), HSP90 and HSP100/105. Most of the proteins are named according to their molecular weight:

HSP70. The 70 Da family of stress proteins is one of the most extensively studied. Included in this family are HSC70 (heat shock cognate, the constitutive form), HSP70 (the inducible form, also referred to as HSP72), and GRP75 (a constitutively expressed glucose-regulated protein found in the endoplasmic reticulum). After a variety of central nervous system (CNS) insults, HSP70 is synthesized at high levels and is present in the cytosol, nucleus and endoplasmic reticulum). Denaturated proteins are thought to serve as a stimulus for induction. These denaturated proteins activate heat shock factors (HSFs) within the cytosol by dissociating other HSPs that are normally bound to HSF (Calabrese et al., 2001a). Freed

HSF is phosphorylated and forms trimers, which enter the nucleus and bind to heat shock elements (HSE) within the promoters of different heat shock genes leading to transcription and synthesis of HSPs. After heat shock, for instance, the synthesis of HSP70 increases to a point where it becomes the most abundant single protein in a cell. Once synthesized, HSP70 binds to denaturated proteins in an ATP-dependent manner. The N-terminal end contains an ATP-binding domain, whereas the C-terminal region contains a substrate-binding domain. Heat shock proteins serve as chaperones that bind to other proteins and regulate their conformation, regulate the protein movement across membranes or through organelles, or regulate the availability of a receptor or activity of an enzyme.

In the nervous system HSPs are induced in a variety of pathological conditions, including cerebral ischemia, neurodegenerative disorders, epilepsy and trauma. Expression of the gene encoding HSPs has been found in various cell populations within the nervous system, including neurons, glia and endothelial cells (Kelly and Yenari, 2002a). HSPs consist of both stress-inducible and constitutive family members. Whether stress proteins are neuroprotective has been the subject of much debate, as it has been speculated that these proteins might be merely an ephiphenomenon unrelated to cell survival. Only recently, however, with the availability of transgenic animals and gene transfer, has it become possible to overexpress the gene encoding HSP70 to test directly the hypothesis that stress proteins protect cells from injury, and it has been demonstrated that overproduction of HSP70 leads to protection in several different models of nervous system injury (Kelly et al., 2002b; Yenari, 2002). Following focal cerebral ischemia, mRNA encoding HSP70 is synthesized in most ischemic cells except in areas of very low blood flow, because of limited ATP levels. HSP70 proteins is produced mainly in endothelial cells, in the core of infarcts in the cells that are most resistant to ischemia, in glial cells at the edges of infarcts and in neurons outside the areas of infarction. It has been suggested that this neuronal expression of HSP70 outside an infarct can be used to define the ischemic penumbras, the zone of protein denaturation in the ischemic areas (Hata et al., 1998). A number of in vitro studies show that both heat shock and HSP overproduction protect CNS cells against both necrosis and apoptosis. Mild heat shock protects neurons against glutamate-mediated toxicity and protects astrocytes against injury produced by lethal acidosis (Narasimham et al., 1996). Transfection of cultured astrocytes with the gene for HSP70 protects them from ischemia or glucose deprivation (Fink et al., 1997). HSP70 has been demonstrated to inhibit caspase-3 activation caused by ceramide, and also affect JUN kinase and p38-kinase activation (Mosser et al., 1997). In addition, HSP70 binds to and modulates the function of BAG-1, the bcl-2 binding protein (McLaughlin et al., 2003), thus modulating some type of apoptosis-related cell death.

A large body of evidence now suggests a correlation between mechanisms of oxidative and/or nitrosative stress and HSP induction. Current opinion hold also the possibility that the heat shock response can exert its protective effects through inhibition of NFkB signaling pathway (Calabrese et al., 2001a). We have demonstrated in astroglial cell cultures that cytokine-induced nitrosative stress is associated with an increased synthesis of HSP70 stress proteins. Increase in HSP70 protein expression was also found after treatment of cells with the NO generating compound sodium nitroprusside (SNP), thus suggesting a role for NO in inducing HSP70 proteins. The molecular mechanisms regulating the NO-induced activation of a heat shock signal seems to involve cellular oxidant/antioxidant balance, mainly represented by the glutathione status and the antioxidant enzymes (McLaughlin et al., 2003; Calabrese et al., 2000b, 2000c).

Ubiquitin is one of the smallest HSPs and is expressed throughout brain in response to ischemia. It is involved in targeting and chaperoning of proteins degraded in proteasomes, which include NFKB, cyclins, HSFs, hypoxia-inducible factor, some apoptosis-related proteins, tumor necrosis factor and erythropoietin receptors (Mayer, 2003).

HSP27 is synthesized mainly in astrocytes in response to ischemic situations or to kainic acid administration. It chaperones cytoskeletal proteins, such as intermediate filaments, actin or glial fibrillary acidic protein following stress in astrocytes. It also protects against Fas-Apo-1, staurosporine, TNF and etoposside-induced apoptotic cell death as well as H₂O₂-induced necrosis (Bechtold and Brown, 2003; Valentim et al., 2003). HSP47 is synthesized mainly in microglia following cerebral ischemia and subarachnoid hemorrhage (Turner et al., 1999).

HSP60, glucose-regulated protein 75 (GRP75) and HSP10 chaperone proteins within mitochondria. GRP75 and GRP78, also called oxygen-regulated proteins (ORPs) are produced by low levels of oxygen and glucose. These protect brain cells against ischemia and seizures *in vivo*, after viral-induced overexpression (Izaki et al., 2001).

HSP32 or heme oxygenase is the rate-limiting enzyme in the production of bilirubin. There are three isoforms of heme oxygenase: HO-1 or inducible isoform, HO-2 or

constitutive isoform and the recently discovered HO-3 (Sahlas et al., 2002; Goldbaum and Richter-Landsberg, 2001; Schipper et al., 1998; Schipper, 2000; Calabrese et al., 2002b; Scapagnini et al., 2002a).

HO as target for neuroprotective strategies in AD pathology

Heme oxygenase is the rate-limiting enzyme in the production of bilirubin. It catalyzes the degradation of heme in a multistep, energy-requiring system. The reaction catalyzed by HO is the α -specific oxidative cleavage of the heme molecule to form equimolar amounts of biliverdin and carbon monoxide (CO). The iron released by HO-1 is bound by ferritin, perhaps via a HO-1 chaperone function (Nath et al., 2000). The redox environment of cytosol and mitochondria is critical to cellular performance and protection against oxidative stress (Drake et al., 2003). Increasing evidence suggests that the HO-1 gene is redox regulated and contains in its promoter region the antioxidant responsive element (ARE), similar to other antioxidant enzymes. Since the expression of heat shock proteins is closely related to that of amyloid precursor protein (APP), heat-shock proteins have been studied in brains of patients with Alzheimer's disease. Significant increases in the levels of HO-1 have been observed in AD brains in association with neurofibrillary tangles (Takeda et al., 2000), and HO-1 mRNA was found to be increased in AD neocortex and cerebral vessels (Premkumar et al., 1995). HO-1 increase was not only in association with neurofibrillary tangles, but also co-localized with senile plaques and glial fibrillary acidic protein-positive astrocytes in AD brains (Takeda et al., 2000). It is conceivable that the dramatic increase in HO-1 in AD may be a direct response to increased free heme associated with neurodegeneration and an attempt to convert the highly damaging heme into the antioxidants biliverdin and bilirubin. Heme oxygenase-1 is rapidly upregulated by oxidative and nitrosative stresses, as well as by glutathione depletion. All these findings have introduced new perspectives in medicine and pharmacology, as molecules activating this defense mechanism appear to be possible candidates for novel cytoprotective strategies (Butterfield et al., 2002).

HO and the therapeutic potential of nutritional antioxidants in AD. Recently, considerable attention has been focused on identifying dietary and medicinal phytochemicals that can inhibit, retard or reverse the multi-stage pathophysiological events underlying AD pathology (Butterfield et al., 2001, 2002a; Calabrese et al., 2000a; Butterfield, 2003). Spice and herbs contain phenolic

V. Calabrese et al.

substances with potent antioxidative and chemopreventive properties (Scapagnini et al., 2002b). The active antioxidant principle in Curcuma longa, a colouring agent and food additive used in Indian culinary preparations, has been identified as curcumin (diferuloylmethane). Due to the presence in its structure of two electrophilic α , β unsaturated carbonyl groups which, by virtue of Michael reaction, can react with nucleophiles such as glutathione, curcumin has the potential to inhibit lipid peroxidation and effectively to intercept and neutralize reactive oxygen and NO-based free radicals (Butterfield and Lauderback, 2002c). This agent is a potent inhibitor of tumor initiation in vivo and possesses antiproliferative activities against tumor cells in vitro (Hayes and McMahon, 2001). Recent epidemiological studies (Ganguli et al., 2000), have raised the possibility that this molecule, as one of the most prevalent nutritional and medicinal compounds used by the Indian population, is responsible for the significantly reduced (4.4-fold) prevalence of AD in India compared to United States. Based on these findings, Lim and colleagues (2000) provided compelling evidence that dietary curcumin given to transgenic APPSw mouse model (Tg2576) for 6 months resulted in a suppression of indices of inflammation and oxidative damage in the brain of this murine model of AD. Furthermore, in a human neuroblastoma cell line it has recently been shown that curcumin inhibits NFkB activation, effectively preventing neuronal cell death Butterfield et al., 2002). Remarkably, recent evidence has demonstrated that curcumin is a potent inducer of HO-1 in vascular endothelial cells (Motterlini et al., 2000a, 2000b). We have also recently demonstrated in astroglial cells the role of caffeic acid phenylethyl ester (CAPE), an active component of propolis, as a novel HO-1 inducer (Scapagnini et al., 2002b). The similarity of CAPE to curcumin is striking because CAPE is also a Michael reaction acceptor, endowed with anti-inflammatory, antioxidant and anticancer effects (Butterfield et al., 2002c). These agents all appear capable of transcriptionally activating a gene battery that includes antioxidant enzymes and heme oxygenase (Dinkova-Kostova et al., 2001). Gene induction occurs through the antioxidant responsive element (ARE) (Hayes and McMahon, 2001). Thus, increased expression of genes regulated by the ARE in cells of the central nervous system may provide protection against oxidative stress.

Vitamin E and AD. Alzheimer's disease (AD) is characterised pathologically by deposition of amyloid β -peptide (A β) in senile (neuritic) plaques, the presence of neurofibrillary tangles, and synapse loss (Katzman and Saitoh, 1991). Oxidative damage in AD brain is extensive,

and $A\beta$ and its sequelae may be associated with this oxidative stress (Butterfield et al., 2001, 2002b). A β causes oxidative damage to and neurotoxicity of neurons (Varadarajan et al., 2000). Vitamin E blocks these effects in vitro (Butterfield et al., 1999, 2002b; Yatin et al., 2000). Plasma from AD and control subjects is low in vitamins E and C, and the level of lipid peroxidation is reportedly inversely correlated with the level of vitamin E (Bourdel-Marchasson et al., 2001). There is a dearth of information about dietary antioxidants on development and progression of neurodegenerative disorders, especially Alzheimer's disease (Halliwell, 2003). Still, trials of vitamin E have been conducted. In one trial, AD patients received 400 IU of vitamin E and 1.0 g of vitamin C daily for one month (Kontush et al., 2001). Plasma and CSF levels of both vitamins E and C were elevated in AD patients relative to baseline, and this increased CSF level of vitamin E was correlated to decreased susceptibility of lipoproteins to in vitro oxidation. A combined approach using the antioxidants vitamins E and C may protect the CNS from oxidative insult, which may reflect the relative stability of the tocopheryl radical and the presence of a reducing agent such as vitamin C to regenerate vitamin E from this radical. Another study of vitamin E and vitamin C supplementation reported that high doses of vitamin E and C supplements may lower the progression of AD symptoms and are protective against vascular dementia (Butterfield et al., 2002a). However, a more recent study suggested that dietary sources rich in vitamin E and C significantly lowered the risk of developing AD relative to supplements alone (Engelhart et al., 2002). One of the largest vitamin E trials in AD is the placebo-controlled, clinical trial of high dose vitamin E (2000 IU per day) in moderate AD patients carried out by the Alzheimer's Disease Cooperative Study (Sano et al., 1997). These large doses of vitamin E, that appeared to pose little risk over an approximately two-year period, were reported to slow the progression of AD. Similar trials involving early AD and mild cognitive impairment subjects are on-going (Grundam, 2000).

However, almost the totality of the vitamin E-based intervention trials are based exclusively on the supplementation of the alpha form of this vitamin while other homologues such as the γ -tocopherol are completely ignored. Emerging evidence on the metabolic fate and biological functions of vitamin E homologues suggests that this could represent a major bias in vitamin E-based intervention trials aimed to prevent oxidative and nitrosative stress in inflammatory and chronic-degenerative disorders (Jiang et al., 2001; Brigelius-Flohé et al., 2002).

The physiologic regulation of vitamin E metabolism make α -tocopherol to be mainly retained and delivered to lipoproteins and tissues while non-alpha vitamers (essentially γ -tocopherols that is abundantly introduced with the diet in western countries) undergo preferential metabolic processing to carboxyethyl hydroxychroman metabolites (CEHC) by shortening of the phytyl chain (Brigelius-Flohé et al., 2002). This metabolic setting results in high plasma ratios of alpha to gamma tocopherol and sustained hepatic degradation of γ -tocopherol to γ -CEHC so that this is the main metabolic product of vitamin E in biological fluids and can be used to mirror the intake of its parental vitamer (Galli et al, 2002; Radosavac et al., 2002; Morinobu et al., 2003). In the brain tissue homogenate the α - to γ -tocopherol ratio is the same than in plasma (Williamson et al., 2002), but in other tissues such as fat and skin the gamma form predominates. This metabolism is also responsible for the lowering effect that a sustained intake of α -tocopherol exerts on the blood levels γ -tocopherol (Brigelius-Flohé et al., 2002; Morton et al., 2002).

These aspects of vitamin E metabolism are particularly important in the light of the recent findings that demonstrated the role of γ -tocopherol as scavenger of reactive nitrogen species (Jiang et al., 2001, 2002; Appenroth et al., 2001), and the contribution that CEHC may give to the antioxidant and anti-inflammatory properties of vitamin E (Betancor-Fernandez et al., 2002; Jiang et al., 2000; Galli et al., 2003). As a consequence, both the gamma homologue of tocopherol and CEHC might play biological roles of relevance in the context of neuroprotection during the different stages of AD. In detail, other than an efficient hydroperoxyl radical scavenger and chain breaker, the gamma configuration of the chroman ring has got the exclusive property to directly scavenge nitric oxidederived species such as peroxynitrite by the formation of a corresponding 5'-nitro- γ -cromanoxyl derivative (Jiang et al., 2001, 2002; Williamson et al., 2002; Appenroth et al., 2001). Again, the gamma configuration of the vitamin E and particularly the γ -CEHC was reported to inhibit both in vitro and in vivo the activity of the pro-inflammatory and pro-oxidant enzyme COX 2 (Jiang et al., 2000, 2002). These specific functions of γ tocopherol and γ -CEHC may thus lead to prevent oxidative and nitrative injury to proteins and lipids in degenerating tissues. Recently, Williamson et al. (2002) have demonstrated the accumulation of 5'-nitro- γ -tocopherol other than 3'-nitro-Tyr in the AD brain and confirmed the superiority of γ -tocopherol vs. α -tocopherol in preventing the peroxynitrite-dependent inactivation of rat brain α -ketoglutarate dehydrogenase as a biomarker of nitrative damage in the AD brain. Gamma tocopherol shows also other important non-antioxidant based functions such as the regulation of cell cycle and apoptotic signalling in proliferating cells (Galli et al., 2003).

Although further studies are required to establish whether the vitamin E could represent an useful tool in the prevention and therapy of neurodegenerative diseases and particularly AD, it is obvious that present intervention studies should be re-examined in the context of these pieces of information. In fact, the disregarded role of γ tocopherol in neuroprotection and antioxidant defence of brain tissues could explain some actually incongruous findings such as the absence of positive effects of supplementing administered as synthetic α -tocopherol opposed to a better outcome obtained in patients to which a diet rich in vitamin E (with all the homologues represented) was administered (Engelhart et al., 2002). At the same time, also the role of CEHC as highly deliverable surrogates of vitamin E could deserve more attention and may explain the positive outcome obtained with large dosages of vitamin E (Sano et al., 1997).

A risk factor for AD is the presence of allele 4 of apolipoprotein E (apoE) (Roses et al., 1996). Synaptosomes from apoE knock-out mice, containing no gene for apoE, show increased susceptibility to oxidative stress induced by A β (Lauderback et al., 2001), while synaptosomes from knock-in mice containing human apoE4 with no mouse background show significantly increased A β induced oxidative stress compared to synaptosomes from human apoE2 or apoE3 knock-in mice (Lauderback et al., 2002). Thus, apoE may serve an antioxidant function, but apoE4 may be less able than apoE2 or apoE3 to do so (Lauderback et al., 2002). This notion was tested using 1month old control and apoE deficient mice. Both received dietary vitamin E for 12 months. Vitamin E-fed animals had better behavioural outcomes of spatial motor activity and decreased levels of lipid peroxidation relative to apoE deficient mice fed a normal diet (Veinbergs et al., 2000). The sum of these studies suggests a decreased risk for and diminished oxidative stress in Alzheimer's disease in persons taking high dose dietary, or perhaps supplemental, vitamin E (and vitamin C to regenerate vitamin E from the tocopherol radical). If proven to be true by additional studies, vitamin E or newer analogues of this antioxidant may prove beneficial in this dementing disorder associated with oxidative stress (Calabrese et al., 2001a; Scapagnini et al., 2002b; Butterfield et al., 2002a).

Conceivably, dietary supplementation with vitamin E or with polyphenolic agents, such as curcumin and its

V. Calabrese et al.

derivatives, can forestall the development of AD, consistent with a major "metabolic" component to this disorder. Nutritional biochemical research is providing optimism that this devastating brain disorder of aging may be significantly delayed and/or modulated.

Acknowledgements

This work was supported, in part, by a grant from the Wellcome Trust (V.C.) and COFIN 2001 (VC, A.M.G.) and by grants from the National Institute of Health (D.A.B.).

References

- Appenroth D, Karge E, Kiessling G, Wechter WJ, Winnefeld K, Fleck C (2001) LLU-alpha, an endogenous metabolite of gamma-tocopherol, is more effective against metal nephrotoxicity in rats than gamma-tocopherol. Toxicol Lett 122: 255–265
- Bechtold DA, Brown IR (2003) Induction of Hsp27 and Hsp32 stress proteins and vimentin in glial cells of the rat hippocampus following hyperthermia. Neurochem Res 28: 1163–1173
- Betancor-Fernandez A, Sies H, Stahl W, Polidori MC (2002) *In vitro* antioxidant activity of 2,5,7,8-tetramethyl-2-(2'-carboxyethyl)-6-hydroxychroman (alpha-CEHC), a vitamin E metabolite. Free Radic Res 36: 915–921
- Bourdel-Marchasson I, Delmas-Beauvieux MC, Peuchant E, Richard-Harston S, Decamps A, Reignier B, Emeriau JP, Rainfray M (2001) Antioxidant defences and oxidative stress markers in erythrocytes and plasma from normally nourished elderly Alzheimer patients. Age Ageing 30: 235–241
- Brigelius-Flohé R, Kelly FJ, Salonen JT, Neuzil J, Zingg JM, Azzi A (2002) The European perspective on vitamin E: current knowledge and future research. Am J Clin Nutr 76: 703–716
- Butterfield DA (2003) Oxidative stress in animal models of accelerated aging, Alzheimer's disease and Huntington's disease. In: Reitz AB, Kordki CP, Choudhary, MI, Rahman AU (eds) Frontiers of medicinal chemistry, Bentham Science Publishers, New York (in press)
- Butterfield DA, Koppal T, Subramaniam R, Yatin S (1999) Vitamin E as an antioxidant/free radical scavenger against amyloid beta-peptide-induced oxidative stress in neocortical synaptosomal membranes and hippocampal neurons in culture: insights into Alzheimer's disease. Rev Neurosci 10: 141–149
- Butterfield DA, Drake J, Pocernich C, Castegna A (2001) Evidence of oxidative damage in Alzheimer's disease brain: central role for amyloid beta-peptide. Trends Mol Med 7: 548–554
- Butterfield DA, Castegna A, Drake J, Scapagnini G, Calabrese V (2002a) Vitamin E and neurodegenerative disorders associated with oxidative stress. Nutr Neurosci 5: 229–239
- Butterfield DA, Castegna A, Pocernich C, Drake J, Scapagnini G, Calabrese V (2002b) Nutritional approaches to combat oxidative stress in Alzheimer's disease. J Nutr Biochem 13: 444–461
- Butterfield DA, Lauderback CM (2002c) Lipid peroxidation and protein oxidation in Alzheimer's disease brain: potential causes and consequences involving amyloid β-peptide-associated free radical oxidative stress. Free Rad Biol Med 32: 1050–1060
- Calabrese V, Renis M, Calderone A, Russo A, Reale S, Barcellona ML, Rizza V (1998) Stress proteins and SH-groups in oxidant-induced cell injury after chronic ethanol administration in rat. Free Rad Biol Med 24: 1159–1167
- Calabrese V, Bates TE, Giuffrida Stella AM (2000a) NO synthase and NO-dependent signal pathways in brain aging and neurodegenerative

- disorders: the role of oxidant/antioxidant balance. Neurochem Res 25: 1315–1341
- Calabrese V, Copani A, Testa D, Ravagna A, Spadaro F, Tendi E, Nicoletti V, Giuffrida Stella AM (2000b) Nitric oxide synthase induction in astroglial cell cultures: Effect on heat shock protein 70 synthesis and oxidant/antioxidant balance. J Neurosci Res 60: 613–622
- Calabrese V, Testa D, Ravagna A, Bates TE, Giuffrida Stella AM (2000c) Hsp70 induction in the brain following ethanol administration in the rat: regulation by glutathione redox state. Biochem Biophys Res Comm 269: 397–400
- Calabrese V, Scapagnini G, Giuffrida Stella AM, Bates TE, Clark JB (2001a) Mitochondrial involvement in brain function and dysfunction: rilevance to aging, neurodegenerative disorders and longevity. Neurochem Research 26: 739–764
- Calabrese V, Scapagnini G, Catalano C, Bates TE, Dinotta F, Micali G, Giuffrida Stella AM (2001b) Induction of heat shock protein synthesis in human skin fibroblasts in response to oxidative stress: regulation by a natural antioxidant from rosemary extract. Int J Tissue React 23: 121–128
- Calabrese V, Scapagnini G, Ravagna A, Giuffrida Stella AM, Butterfield DA (2002a) Molecular chaperones and their roles in neural cell differentiation. Dev Neurosci 24: 1–13
- Calabrese V, Scapagnini G, Ravagna A, Fariello RG, Giuffrida Stella AM, Abraham NG (2002b) Regional distribution of heme oxygenase, HSP70, and glutathione in brain: relevance for endogenous oxidant/antioxidant balance and stress tolerance. J Neurosci Res 68: 65–75
- Dinkova-Kostova AT, Massiah MA, Bozak RE, Hicks RJ, Talalay P (2001) Potency of Michael reaction acceptors as inducers of enzymes that protect against carcinogenesis depends on their reactivity with sulfhydryl groups. Proc Natl Acad Sci USA 98: 3404–3409
- Drake J, Sultana R, Aksenova M, Calabrese V, Butterfield DA (2003) Elevation of mitochondrial glutathione by γ -glutamylcysteine ethyl ester protects mitochondria against peroxynitrite-induced oxidative stress. J Neurosci Res (in press)
- Engelhart MJ, Geerlings MI, Ruitenberg A, van Swieten JC, Hofman A, Witteman JCM, Breteler MMB (2002) Dietary intake of antioxidants and risk of Alzheimer's disease. JAMA 287: 3223–3227
- Fink SL, Chang LK, Ho DY, Sapolsky RM (1997) Defective herpes simplex virus vectors expressing the rat brain stress-inducible heat shock protein 72 protect cultured neurons from severe heat shock. J Neurochem 68: 961–969
- Galli F, Lee R, Dunster C, Kelly FJ (2002) Gas chromatography mass spectrometry analysis of carboxyethyl-hydroxychroman metabolites of alpha- and gamma-tocopherol in human plasma. Free Rad Biol Med 32: 333–340
- Galli F, Stabile AM, Betti M, Piroddi M, Floridi A, Azzi A (2003) The γ -tocopherol metabolite γ -carboxyethyl hydroxychroman acid inhibits prostate cancer cell (PC-3) proliferation: a preliminary characterization. Arch Biochem Biophys (in press)
- Ganguli M, Chandra V, Kamboh MI, Johnston JM, Dodge HH, Thelma BK, Juyal RC, Pandav R, Belle SH, DeKosky ST (2000) Apolipoprotein E polymorphism and Alzheimer disease: The Indo-US Cross-National Dementia Study. Arch Neurol 57: 824–830
- Goldbaum O, Richter-Landsberg C (2001) Stress proteins in oligodendrocytes: differential effects of heat shock and oxidative stress. J Neurochem 78: 1233–1242
- Goodman R, Blank M (2002) Insights into electromagnetic interaction mechanisms. J Cell Physiol 192: 16–22
- Grundman M (2000) Vitamin E and Alzheimer disease: the basis for additional clinical trials. Am J Clin Nutr 71: 630S-636S
- Halliwell B (2001) Role of free radicals in the neurodegenerative diseases: therapeutic implications for antioxidant treatment. Drugs and Aging 18: 685–716
- Hata R, Gass P, Mies G, Wiessner C, Hossmann KA (1998) Attenuated cfos mRNA induction after middle cerebral artery occlusion in CREB

- knockout mice does not modulate focal ischemic injury. J Cereb Blood Flow Metab 18: 1325–1335
- Hayes JD, McMahon M (2001) Molecular basis for the contribution of the antioxidant responsive element to cancer chemoprevention. Cancer Lett 174: 103–1013
- Izaki K, Kinouchi H, Watanabe K, Owada Y, Okubo A, Itoh H, Kondo H, Tashima Y, Tamura S, Yoshimoto T, Mizoi K (2001) Induction of mitochondrial heat shock protein 60 and 10 mRNAs following transient focal cerebral ischemia in the rat. Brain Res Mol Brain Res 31: 14–25
- Jiang Q, Elson-Schwab I, Courtemanche C, Ames BN (2000) Gammatocopherol and its major metabolite, in contrast to alpha-tocopherol, inhibit cyclooxygenase activity in macrophages and epithelial cells. Proc Natl Acad Sci USA 97: 11494–11499
- Jiang Q, Christen S, Shigenaga MK, Ames BN (2001) gamma-tocopherol, the major form of vitamin E in the US diet, deserves more attention. Am J Clin Nutr 74: 714–722
- Jiang Q, Lykkesfeldt J, Shigenaga MK, Shigeno ET, Christen S, Ames BN (2002) G-tocopherol supplementation inhibits protein nitration and ascorbate oxidation in rats with inflammation. Free Rad Biol Med 33: 1534–1542
- Katzman R, Saitoh T (1991) Advances in Alzheimer's disease. FASEB J 5: 278–286
- Kelly S, Yenari MA (2002a) Neuroprotection: heat shock proteins. Curr Med Res Opin 18: 55–60
- Kelly S, Zhang ZJ, Zhao H, Xu L, Giffard RG, Sapolsky RM, Yenari MA, Steinberg GK (2002b) Gene transfer of HSP72 protects cornu ammonis 1 region of the hippocampus neurons from global ischemia: influence of Bcl-2. Ann Neurol 52: 160–167
- Kontush A, Mann U, Arlt S, Ujeyl A, Luhrs C, Muller-Thomsen T, Beisiegel U (2001) Influence of vitamin E and C supplementation on lipoprotein oxidation in patients with Alzheimer's disease. Free Rad Biol Med 31: 345–354
- Lauderback CM, Hackett JM, Keller JN, Varadarajan S, Szweda L, Kindy M, Markesbery WR, Butterfield DA (2001) Vulnerability of synaptosomes from apoE knock-out mice to structural and oxidative modifications induced by $A\beta(1-40)$: implications for Alzheimer's disease. Biochemistry 40: 2548–2554
- Lauderback CM, Kanski J, Hackett JM, Maeda N, Kindy MS, Butterfield DA (2002) Apolipoprotein E modulates Alzheimer's A β (1–42)-induced oxidative damage to synaptosomes in an allele-specific manner. Brain Res 924: 90–97
- Lim GP, Yang F, Chu T, Chen P, Beech W, Teter B, Tran T, Ubeda O, Hsiao Ashe K, Frautschi SA, Cole GM (2000) Ibuprofen suppresses plaque pathology and inflammation in a mouse model for Alzheimer's disease. J Neurosci 20: 5709–5714
- Mattson MP, Duan W, Chan SL, Cheng A, Haughey N, Gary DS, Guo Z, Lee J, Furukawa K (2002) Neuroprotective and neurorestorative signal transduction mechanisms in brain aging: modification by genes, diet and behavior. Neurobiol Aging 23: 695–705
- Mayer RJ (2003) From neurodegeneration to neurohomeostasis: the role of ubiquitin. Drug News Perspect 16: 103–108
- McLaughlin B, Hartnett KA, Erhardt JA, Legos JJ, White RF, Barone FC, Aizenman E (2003) Caspase 3 activation is essential for neuroprotection in preconditioning. Proc Natl Acad Sci USA 100: 715–720
- Misseri R, Shaw MB, Gearhart JP, Meldrum DR (2003) Liposomal delivery of heat shock protein 72 into renal tubular cells blocks nuclear factor-kappaB activation, tumor necrosis factor-alpha production, and subsequent ischemia-induced apoptosis. Circ Res 92: 293–299
- Morinobu T, Yoshikawa S, Hamamura K, Tamai H (2003) Measurement of vitamin E metabolites by high-performance liquid chromatography during high-dose administration of alpha-tocopherol. Eur J Clin Nutr 57: 410–414
- Morton LW, Ward NC, Croft KD, Puddey IB (2002) Evidence for the nitration of gamma-tocopherol in vivo: 5-nitro-gamma-tocopherol is

- elevated in the plasma of subjects with coronary heart disease. Biochem J 364: 625–628
- Mosser DD, Caron AW, Bourget L, Denis-Larose (1997) Role of the human heat shock protein hsp70 in protection against stress-induced apoptosis. Mol Cell Biol 17: 5317–5327
- Motterlini R, Foresti R, Bassi R, Green CJ (2000a) Curcumin, an antioxidant and anti-inflammatory agent, induces heme oxygenase-1 and protects endothelial cells against oxidative stress. Free Radic Biol Med 28: 1303–1312
- Motterlini R, Foresti R, Bassi R, Calabrese V, Clark JE, Green CJ (2000b) Endothelial Heme oxygenase-1 induction by hypoxia: modulation by inducible nitric oxide synthase (iNOS) and S-nitrosothiols. J Biol Chem 275: 13613–13620
- Narasimhan P, Swanson RA, Sagar SM, Sharp FR (1996) Astrocyte survival and HSP70 heat shock protein induction following heat shock and acidosis. Glia 17: 147–159
- Nath KA, Haggard JJ, Croatt AJ, Grande JP, Poss KD, Alam J (2000) The indispensability of heme oxygenase-1 in protecting against acute heme protein-induced toxicity in vivo. Am J Pathol 156: 1527–1535
- Premkumar DR, Smith MA, Richey PL, Petersen RB, Castellani R, Kutty RK, Wiggert B, Perry G, Kalaria RN (1995) Induction of heme oxygenase-1 mRNA and protein in neocortex and cerebral vessels in Alzheimer's disease. J Neurochem 65: 1399–1402
- Radosavac D, Graf P, Polidori MC, Sies H, Stahl W (2002) Tocopherol metabolites 2, 5, 7, 8-tetramethyl-2-(2'-carboxyethyl)-6-hydroxychroman (alpha-CEHC) and 2, 7, 8-trimethyl-2-(2'-carboxyethyl)-6-hydroxychroman (gamma-CEHC) in human serum after a single dose of natural vitamin E. Eur J Nutr 41: 119–124
- Ropeleski MJ, Tang J, Walsh-Reitz MM, Musch MW, Chang EB (2003) Interleukin-11-induced heat shock protein 25 confers intestinal epithelial-specific cytoprotection from oxidant stress. Gastroenterology 124: 1358–1368
- Roses AD (1996) Apolipoprotein E alleles as risk factors in Alzheimer's disease. Ann Rev Med 47: 387–400
- Sahlas DJ, Liberman A, Schipper HM (2002) Role of heme oxygenase-1 in the biogenesis of corpora amylacea. Biogerontology 3: 223–231
- Sano M, Ernesto C, Thomas RG, Klauber MR, Schafer K, Grundman M, Woodbury P, Growdon J, Cotman CW, Pfeiffer E, Schneider LS, Thal LJ (1997) A controlled trial of selegiline, alpha-tocopherol, or both as treatment for Alzheimer's disease. The Alzheimer's Disease Cooperative Study. N Engl J Med 336: 1216–1222
- Scapagnini G, D'Agata V, Calabrese V, Pascal A, Colombrita C, Alkon D, Cavallaro S (2002a) Gene expression profiles of heme oxygenase isoforms in the rat brain. Brain Res 954: 51–59
- Scapagnini G, Foresti R, Calabrese V, Giuffrida Stella AM, Green CJ, Motterlini R (2002b) Caffeic acid phenethyl ester and curcumin: a novel class of heme oxygenase-1 inducers. Mol Pharmacol 61: 554–561
- Schipper HM (2000) Heme oxygenase-1: role in brain aging and neurodegeneration. Exp Gerontol 35: 821–830
- Schipper HM, Liberman A, Stopa EG (1998) Neural Heme oxygenase-1 expression in idiopathic Parkinson's disease. Exp Neurol 150: 60–68
- Takeda A, Perry G, Abraham NG, Dwyer BE, Kutty RK, Laitinen JT, Petersen RB, Smith MA (2000) Overexpression of heme oxygenase in neuronal cells, the possible interaction with Tau. J Biol Chem 275: 5395–5399
- Turner CP, Panter SS, Sharp FR (1999) Anti-oxidants prevent focal rat brain injury as assessed by induction of heat shock proteins (HSP70, HO-1/HSP32, HSP47) following subarachnoid injections of lysed blood. Brain Res Mol Brain Res 65: 7–102
- Valentim LM, Rodnight R, Geyer AB, Horn AP, Tavares A, Cimarosti H, Netto CA, Salbego CG (2003) Changes in heat shock protein 27 phosphorylation and immunocontent in response to preconditioning

- to oxygen and glucose deprivation in organotypic hippocampal cultures. Neuroscience 118: 379-386
- Varadarajan S, Yatin S, Aksenova M, Butterfield DA (2000) Review: Alzheimer's amyloid beta-peptide-associated free radical oxidative stress and neurotoxicity. J Struct Biol 130: 184–208
- Veinbergs I, Mallory M, Sagara Y, Masliah E (2000) Vitamin E supplementation prevents spatial learning deficits and dendritic alterations in aged apolipoprotein E-deficient mice. Eur J Neuroscience 12: 4541–4546
- Wheeler DS, Dunsmore KE, Wong HR (2003) Intracellular delivery of HSP70 using HIV-1 Tat protein transduction domain. Biochem Biophys Res Commun 301: 54–59
- Williamson KS, Gabbita SP, Mou S, West M, Pye QN, Markesbery WR, Cooney RV, Grammas P, Reimann-Philipp U, Floyd RA, Hensley K

- (2002) The nitration product 5-nitro-g-tocopherol is increased in the Alzheimer brain. Nitric Oxide Biol Chem 6: 221–227
- Yatin SM, Varadarajan S, Butterfield DA (2000) Vitamin E prevents Alzheimer's amyloid β -peptide (1–42)-induced protein oxidation and reactive oxygen species formation. J Alzheimer's Dis 2: 123–131
- Yenari MA (2002) Heat shock proteins and neuroprotection. Adv Exp Med Biol 513: 281–299

Authors' address: Prof. Vittorio Calabrese, Section of Biochemistry and Molecular Biology, Department of Chemistry, Faculty of Medicine, University of Catania, Viale Andrea Doria 6, 95100 Catania, Italy, Fax: 0039-095-580138, E-mail: calabres@mbox.unict.it